# MICROBIAL TRANSFORMATION OF ANTIBIOTICS

## II. ADDITIONAL TRANSFORMATION PRODUCTS OF MARIDOMYCIN III

MOTOO SHIBATA, MASARU UYEDA and SHINYA MORI

Laboratory of Medicinal Microbiology, Faculty of Pharmaceutical Sciences, Kumamoto University, Kumamoto City, 862 Japan

(Received for publication April 19, 1976)

Streptomyces sp. strain No. K-245 was found to transform maridomycin III into four derivatives ( $A_1$ ,  $A_2$ ,  $A_3$  and  $A_4$ ) in addition to the transformation products reported previously. Isolation of the main product  $A_1$  was carried out by column chromatography on silica gel developed with CHCl<sub>8</sub>- MeOH (19: 1). From the partial investigation of the structure of  $A_1$ , it proved to have a C 18-aldehyde group and C 4"-propionyl group but no antimicrobial activity. The relationships between A group's derivatives and known derivatives of maridomycin III are also discussed.

The studies on microbial transformation of antibiotics have been carried out in order to improve the antimicrobial activity of existing ones, to make them less-toxic or more effective against resistant strains. In the studies on microbial transformation of 16-membered macrolide antibiotics, 4"deacylation<sup>1~4</sup>), 3"-hydroxylation<sup>5,6</sup>), 14-hydroxylation<sup>7</sup>) and reduction of aldehyde<sup>1,8,0</sup>) have been reported.

In our previous paper<sup>1)</sup>, *Streptomyces lavendulae* strain No. K-122 was shown to transform maridomycin (MDM) III<sup>10,11)</sup> into three derivatives. Of these three transformation products, 18-dihydro-MDM III and 4"-depropionyl-MDM III were identical with those previously reported by NAKAHAMA *et al.*<sup>9,2,8)</sup>, whereas 18-dihydro-4"-depropionyl-MDM III was a new transformation product. The relationship between MDM III and its transformation products was considered to be as shown in Fig. 1; that is, 4"-depropionyl-MDM III was formed by deacylation of MDM III, and 18-dihydro-MDM III by reduction of MDM III. 18-Dihydro-4"-depropionyl-MDM III was formed either from 4"depropionyl-MDM III by reduction or from 18-dihydro-MDM III by deacylation.

Streptomyces sp. strain No. K-245 was found to transform MDM III into four derivatives in addition to the transformation products described above. This paper deals with the isolation of the

transformation products formed by strain No. K-245 and the relationship between MDM III and these derivatives.

#### Materials and Methods

### Antibiotics

Maridomycin (MDM) III (cf. Fig. 1), and 4''depropionyl MDM III were kindly supplied by Takeda Chemical Industries, Ltd. 18-Dihydro-MDM, 4''-depropionyl-MDM III (in part) and 18-dihydro-4''-depropionyl-MDM III were prepared by the transformation of MDM III by *S. layendulae* strain No. K-122.<sup>1)</sup> Fig. 1. Transformation of MDM III into its derivatives by *Streptomyces lavendulae* strain No. K-122.



### THE JOURNAL OF ANTIBIOTICS

# VOL. XXIX NO. 8

## Cultural Conditions

Transformation was carried out in  $20 \sim 25$  ml of a medium in 200-ml Erlenmeyer flasks. The seed culture medium containing 2% glucose, 3% soluble starch, 1% soybean flour, 1% corn steep liquor, 0.5% Polypepton, 0.3% NaCl and 0.5% CaCO<sub>8</sub> (pH 7.0), was inoculated with spores from the slant culture. *Streptomyces* sp. strain No. K-245 was grown for 24 hours at 28°C on a rotary shaker (180 rpm, 5 cm radius).

This culture was used to inoculate the transformation medium (GC medium) containing 5% glucose, 0.5% Polypepton, 2% cornsteep liquor, 0.3% NaCl and 0.5% CaCO<sub>3</sub> (pH7.0). After 48 hours of cultivation a conc. solution of MDM III in methanol was added to the culture (final concentration of MDM III:  $1 \sim 2$  mg/ml medium), and then the fermentation was continued for an additional 48 hours.

Thin-layer Chromatography (TLC)

The filtered broth was extracted with  $CHCl_3$  at pH 8.2 and the solvent layer was chromatographed on silica gel G (Merck) plates using  $CHCl_3$ -MeOH (17: 3, v/v) as solvent. The position of the products was detected by heating the plates after spraying with 10% H<sub>2</sub>SO<sub>4</sub>.

Detection of O-Propionylmycarose<sup>12,13)</sup>

The samples were hydrolyzed with  $0.5 \times HCl$  for 24 hours at room temperature, extracted with ether at pH 4.0, and the solvent layer was chromatographed (TLC) using 1-butanol - acetic acid - water (4: 1: 5, upper layer). O-Acylmycarose was detected by heating for 10 minutes at 90°C after spraying with vanillin-perchloric acid reagent<sup>14</sup>).

#### Results

# Transformation of MDM III by Growing and Washed Cells

Streptomyces sp. strain No. K-245 transformed MDM III into four derivatives in GC medium, whereas the other three derivatives formed by *S. lavendulae*<sup>1)</sup> were also detected in glucose-bouillon and similar media. Typical thin-layer chromatograms of the products produced in GC medium are shown in Fig. 2. At first this strain was found to produce an unknown derivative, designated temporarily as A in Fig. 2(a). By changing the solvent system to CHCl<sub>3</sub>-MeOH (17: 3), A proved to consist of four derivatives as shown in Fig. 2(b) and designated as A<sub>1</sub>, A<sub>2</sub>, A<sub>3</sub> and A<sub>4</sub> in the order of their decreasing Rf values on TLC. The same results were obtained with washed cells.

Isolation of Main Transformation Product A1

Filtered broth was concentrated *in vacuo* by azeotropic distillation with 1-butanol, crude A complex (consisting of  $A_1$ ,  $A_2$ ,  $A_3$  and  $A_4$ ) was extracted with CHCl<sub>3</sub> at pH 8.2, and concentrated *in vacuo*. As shown in Fig. 3, crude  $A_1$  was dissolved in CHCl<sub>3</sub>, decolorized by activated charcoal and concentrated

*in vacuo.* Purified  $A_1$  was precipitated by the addition of *n*-hexane. The yield of  $A_1$  from crude A complex was shown in Table 1. The mobility of purified  $A_1$  and other derivatives was examined by TLC with results as shown in Table 2.

# Physicochemical Properties of A1

 $A_1$  is a white-colored, basic material and its melting point is 134~135°C. It showed only end absorption and the results of elementary analysis for  $A_1$  were as follows: C 58.36, 58.39; H 8.21, 8.18; N 1.20, 1.17 (%). Its molecular Fig. 2. Typical thin-layer chromatograms of MDM III transformation products by *Streptomyces* sp. strain No. K-245.

1: MDM III, 2: 18-dihydro-MDM III, 3: 4"depropionyl-MDM III, 4: 18-dihydro-4"-depropionyl-MDM III, A: A complex.



Fig. 3.	Purification	procedure	of	main	transforma-	
tion 1	product A <sub>1</sub> .					

С	ulture filtrate
	adjustment of pH to 8.2
	extraction with CHCl <sub>3</sub>
C	HCl <sub>3</sub> layer
	concentration in vacuo
C	rude A complex
	silica gel chromatography
	$(CHCl_{8} - MeOH, 19:1)$
A	1 fraction
	concentration <i>in vacuo</i> , decolorization with activated charcoal in CHCl.
С	HCl <sub>3</sub> layer
	concentration <i>in vacuo</i> precipitation with n-hexane

Purified A<sub>1</sub>

weight is  $860 \sim 890$  by vapor pressure osmometry in CHCl<sub>3</sub>. It is positive in the ELSON-MORGAN test and negative in the anthrone, FEHLING, erythromycin and carbomycin tests. Color reactions of A<sub>1</sub> and MDM III were similar.

Since  $A_1$  degraded during mass spectrometry,  $A_1$  was acetylated by acetic anhydride in pyridine according to the method described by KINUMAKI *et al.*<sup>15)</sup> The NMR spectrum of acetylated  $A_1$  is shown in Fig. 4.\* A proton at

Table 1. Yields of  $A_1$  from crude A complex.

	Weight (mg)	Yield (%)
Crude A complex	720	100
A <sub>1</sub> fraction	145.8	20.3
A <sub>1</sub> after decolorization	123.5	17.2
Purified A <sub>1</sub>	113.1	15.7

Table 2. Mobility of components  $A_1 \sim A_4$  by thinlayer chromatography.

C	Rf in the indicated solvent system					
Component	(1)	(2)	(3)	(4)		
A1	0.53	0.12	0.62	0.28		
$A_2$	0.41	0.10	0.62	0.20		
$A_3$	0.21	0.05	0.50	0.12		
$A_4$	0.13	0.02	0.50	0.03		

Solvent system:

(1) CHCl<sub>3</sub> - MeOH (17:3)

(2)  $C_6H_6 - Me_2CO$  (1:2)

(3) 1-BuOH - AcOH -  $H_2O$  (3:1:1)

(4)  $C_6H_6$  - MeOH (4:1)

 $\delta$  3.53 as C 4-methoxy group and a proton at  $\delta$  2.53 as a dimethylamino group of C 3 in mycaminose were still seen, which means there is no transformation on these positions. These protons were also seen in the NMR spectrum of free A<sub>1</sub>. Moreover, simple depropionylation at C 4 in mycarose did not occur since the fragment ion peak at O-propionylmycarose (*m/e* 201) was seen in the mass spectrum of acetylated A<sub>1</sub>. This fact is in good agreement with the result in the detection of O-propionylmycarose described below. Examination of the final structure of A<sub>1</sub> is now in progress and will be reported in a separate paper.

> Transformation of MDM III into  $A_1$ ,  $A_2$ ,  $A_3$  and  $A_4$ and their Relationship to the Known Derivatives

Transformations of MDM III, 18-dihydro-MDM III, 4''-depropionyl-MDM III, 18-dihydro-4''depropionyl-MDM III or A<sub>1</sub>, by *Streptomyces* K-245 were examined. A<sub>1</sub> and A<sub>8</sub> were formed from MDM III, A<sub>2</sub> and A<sub>4</sub> from 18-dihydro-MDM III, A<sub>8</sub> from 4''-depropionyl-MDM III, A<sub>4</sub> from 18dihydro-4''-depropionyl-MDM III and that A<sub>8</sub> was also formed from A<sub>1</sub>.

Moreover, O-propionylmycarose was detected in both MDM III and  $A_1$  and not in 4"-depropionyl-MDM III and  $A_8$ . This result suggests that the transformation reaction of  $A_1$  into  $A_8$  is a deacylation reaction which was observed in the transformation of MDM III into 4"-depropionyl-MDM III.

From these results, the transformation pathway was diagrammed as shown in Fig. 5. In addition to the inner pathway previously reported<sup>1)</sup>,  $A_1$ ,  $A_2$ ,  $A_3$  or  $A_4$  was formed from MDM III, 18-dihydro-MDM III, 4"-depropionyl-MDM III or 18- dihydro-4"-depropionyl-MDM III, respectively, *via* trans-

<sup>\*</sup> A proton at  $\delta$  9.62 which was recognized as an aldehyde proton in MDM III was still present.

formation at a common position.  $A_1$  itself was transformed into  $A_2$ ,  $A_3$  and  $A_4$  by reduction, deacylation or both in the same pathway as the inner one.

### Discussion

*Streptomyces* sp. strain No. K-245 transformed MDM III into four derivatives. These derivatives were thought to be trans-

formed *via* common transformation from MDM III, 18-dihydro-, 4''-depropionyl-, or 18-dihydro-4''-depropionyl-MDM III, respectively.

Although the antimicrobial spectrum is not shown in this paper, main product  $A_1$ , for example, lost all activity (retained 0.1% or less of activity of MDM III). It is interesting in the relationship between structure and antimicrobial activity that  $A_1$  still has a C 18-aldehyde group which is one of the essential groups for antimicrobial activity of the macrolide antibiotics<sup>1,3,16,17)</sup>.

Fig. 4. NMR spectrum of acetylated A<sub>1</sub>.



Fig. 5. Transformation of MDM III into its derivatives by *Streptomyces* K-245.



A-group's derivatives may be unknown or new microbial transformation products since the position transformed is in the other position than C 18-aldehyde and 4"-propionyl group. The final structure will be reported in the future.

#### Acknowledgment

We wish to thank Takeda Chemical Industries, Ltd. for the supply of maridomycin and 4"-depropionyl-MDM III and for mass- and NMR-analyses. We also thank Miss M. MORITA for her technical assistance.

#### References

- 1) SHIBATA, M.; M. UYEDA & S. MORI: Microbial transformation of antibiotics. I. Isolation and characterization of the transformation products of maridomycin III. J. Antibiotics 28: 434~441, 1975
- NAKAHAMA, K.; M. IZAWA, T. MUROI, T. KISHI, M. UCHIDA & S. IGARASI: Microbial conversion of antibiotics. I. Deacylation of maridomycin by bacteria. J. Antibiotics 27: 425~432, 1974
- NAKAHAMA, K.; T. KISHI & S. IGARASI: Microbial conversion of antibiotics. II. Deacylation of maridomycin by actinomycetes. J. Antibiotics 27: 487~488, 1974
- MUROI, M.; M. IZAWA & T. KISHI: Maridomycin, a new macrolide antibiotic. VIII. Isolation and structure of metabolites of 9-propionylmaridomycin. J. Antibiotics 27: 449~459, 1974
- NAKAHAMA, K.; T. KISHI & S. IGARASI: Microbial conversion of antibiotics. III. Hydroxylation of maridomycin I and josamycin. J. Antibiotics 27: 433~441, 1974
- TACHIBANA, A.; M. SHIBATA, F. KUMAGAI, K. MORIYAMA, K. YANO & T. OSONO: Studies on josamycin propionate. II. Distribution and metabolism. Jap. J. Antibiotics 26: 122~129, 1973
- INOUYE, S.; T. SHOMURA, T. TSURUOKA, S. OMOTO, T. NIIDA & K. UMEZAWA: Isolation and structure of two metabolites of a macrolide antibiotic SF-837 substance. Chem. Pharm. Bull. (Tokyo) 20: 2366~2371, 1972
- FELDMAN, L. I.; I. K. DILL, G. E. HOLMLUND, H. A. WHALEY, E. L. PATTERSON & N. BOHONOS: Microbiological transformation of macrolide antibiotics. Antimicr. Agents & Chemoth.-1963: 54~57, 1964
- 9) NAKAHAMA, K. & S. IGARASI: Microbial conversion of antibiotics. IV. Reduction of maridomycin.

J. Antibiotics 27: 605~609, 1974

- MUROI, M.; M. IZAWA & T. KISHI: Structure of maridomycin I, III, IV, V and VI, macrolide antibiotics. Experientia 28: 129~131, 1972
- ONO, H.; T. HASEGAWA, E. HIGASHIDE & M. SHIBATA: Maridomycin, a new macrolide antibiotic. I. Taxonomy and fermentation. J. Antibiotics 26: 191~198, 1973
- WATANABE, T.; T. FUJII & K. SATAKE: 4-O-Acetyl mycarose, a new O-acetyl sugar obtained from leucomycin minor components. J. Biochem. 50: 197~201, 1961
- WATANABE, T.; H. NISHIDA & K. SATAKE: Studies on leucomycin V. Isolation of mycarose-4-isovalerate from leucomycin A<sub>1</sub>. Chem. Pharm. Bull. 34: 1285~1288, 1961
- MACLENNAN, A. P. & H. A. RANDALL: Detection and identification of deoxysugars on paper chromatograms. Anal. Chem. 31: 2020~2022, 1950
- 15) KINUMAKI, A.; I. TAKAMORI, Y. SUGAWARA, M. SUZUKI & T. OKUDA: Studies on the macrolide antibiotics YL-704 complex. III. The structure of new macrolide antibiotics YL-704 A<sub>1</sub> and B<sub>1</sub>. J. Antibiotics 27: 107~116, 1974
- 16) ADAMSKI, R. J.; H. HEYMANN, S. G. GEFTIC & S. S. BARKULIS: Preparation and antibacterial activity of some spiramycin derivatives. J. Med. Chem. 9: 932~934, 1966
- 17) OMURA, S. & M. TISHLER: Relationship of structures and microbiological activities of the 16-membered macrolides. J. Med. Chem. 15: 1011~1015, 1972